SOP Title: Agilent RNA 6000 Pico Quality Control	Version 1.1, Page 1	
Authorized Signature:	Issuing Date: 02.09.11	
	Last Revision Date: 11.30.11 - SV	
Staff Able to Perform Procedure: Research Technician and higher		
Principle of the Method:		
This protocol describes the initial quality and quantification control steps when total RNA is received by		
the UB Next-Generation Sequencing and expression Analysis Core.		
Sample Type: total RNA sample		
Equipment Requirements:		
o Agilent 2100 Bioanalyzer (Agilent)		
o Bioanalyzer Chip Vortex (IKA)		
O VORTEX (VVVR)		
o Microcentriluge (VWR)		
O IIIIei (IIIou) Descente & Material Dequiremente:		
Quant-IT Ribogreen Assay (Invitrogen)		
o PMMA Cuvettes (Fischer)		
 D FINIMA Cuvcues (Fischer) D RNAse, free centrifuge tubes 15ml 200 µl (USA Scient) 	o RNAse- free centrifuge tubes 15ml 200 ul (USA Scientific)	
• Pipette Man 2ul 20ul 200ul 1000ul (Rainin)		
• Filter Pinette Tips (Rainin)		
Sample		
o Wearing gloves, place total RNA samples at -80°C in labeled 5x5 Crvo freezer box		
Prenaring the Gel		
• Pipette 550 ul of RNA 6000 Pico gel matrix (red) into a spin filter		
\circ Centrifuge at 1500 g for 10 minutes at room temperature		
• Aliquot 65 ul filtered gel into 0.5 ml microfuge tubes. Use filtered gel within 4 weeks		
Prenaring the Gel-Dye Mix		
• Allow RNA 6000 Pico dve concentrate (blue) to equilibrate to room temperature for 30 minutes		
• Vortex RNA 6000 Pico dye concentrate (blue) for 10 se	• Vortex PNA 6000 Pico dye concentrate (blue) for 10 seconds spin down, and add Lul of dye into	
a 65 ul aliquot of filtered gel		
ο Vortex solution well		
\circ Spin tube at 13000 g for 10 minutes at room temperatur.	e Use prepared Gel-Dye mix within one	
dav	e. Ose prepared der Dye link within one	
Loading the Gel-Dye Mix		
o Adjust the syringe clip so it is at the highest position.		
• Put a new RNA 6000 chip on the chip priming station.		
\circ Pipette 9.0 µl of gel-dye mix in the well marked \widehat{G}		
• Make sure that the plunger is positioned at 1 ml and then close the chip priming station.		
• Press plunger until it is held by the clip.	• Press plunger until it is held by the clip.	
• Wait exactly 30 seconds then release clip.	o Wait exactly 30 seconds then release clip.	
o Wait for 5 seconds. Slowly pull back plunger to 1 ml position.		
o Open the chip priming station and pipette 9.0 µl of gel-	• Open the chip priming station and pipette 9.0 μ of gel-dye mix in the wells marked G	
o Discard the remaining gel-dye mix.		
Loading the RNA 6000 Pico Conditioning Solution and Markers		
ο Pipette 9.0 μl of the RNA 6000 Pico Condition Solution (white) in the well marked CS.		
• Pipette 5 µl of marker (green) in all 11 sample wells and the ladder well.		
Loading the Diluted Ladder and Samples		
\circ Pipette 1 µl of the heat denatured and aliquoted ladder i	in the well marked ladder.	

- ο In each of the 11 sample wells pipette 1 μl of sample (used wells) or 1 μl of RNA 6000 Pico Marker (green) (unused wells).
- Put the chip horizontally in the adapter and vortex for 1 minute at the indicated setting (2400 rpm).
- o Run the chip in the Agilent 2100 bioanalyzer within 5 minutes.